Please replace the paragraph beginning on page 82, line 8 with the following rewritten paragraph:

-- There is no experimental work on introns in *Candida*. So we selected one possible candidate, the very small intron (mini-intron) from the peptide transporter gene (Basrai et al 1995). This was amplified by PCR and inserted into the URA3 gene in both the forward and backward direction (Figure 74). The forward was a control to make sure the peptide transporter intron would splice. As expected, it did. --

Please replace the paragraph beginning on page 83, line 6 with the following rewritten paragraph:

-- We have now mounted this URA3/inverted intron element onto a retrotransposon plasmid putting the element into a (synthetic) Nsi1 site at the 3′ end of the coding sequence. We have also added an ADE2 element between the right LTR and the *Candida* ARS (CARS). This is summarised in Figure 75. --

Please replace the paragraph beginning on page 84, line 1 with the following rewritten paragraph:

-- The plasmid is quite large and therefore not that easy to work with but it has been completed. The plasmid has been transformed into two ADE2$^-$URA$^-$ strains, one carrying a URA3 point mutation and the other a URA3 deletion (a small deletion) (Figure 76). ADE2$^+$ transformants were selected and grown at 37°C to encourage retrotransposition. Cultures were then plated on minimal medium + adenine. The plasmid is lost under these conditions and only URA$^+$ variants (retrotranspositions?) can grow. Both strains gave URA$^+$ derivatives. The URA$^-$ point mutation is reasonably stable and the URA$^-$ deletion completely so. We, therefore, are sure that these URA$^+$ variants are not revertants. They are, we believe, a mixture of retrotransposition and gene conversion. There is very little literature on gene conversion in *Candida*. --

Please replace the paragraph beginning on page 86, line 1 with the following rewritten paragraph:

-- These should only give a product following a retrotransposition event since the intron must be removed before primer i) will work (Figure 77). --

Please add the following paragraph on page 22, line 8 in the Brief Description of the Drawings:

-- Figure 73 is a schematic representation of an inverted intron inserted within the reporter gene, URA3. A) The transcript is not able to code for the URA3 gene product because the intron cannot be removed (it is in backwards). B) The transcript before and after splicing from Retrotransposon promoter (pRet). The transcript is not able to code for the URA3 gene product because, although the intron can be removed (processed or spliced), the URA3 sequence is backwards. C) Reverse transcriptase/integrase functions of the retrotransposon may act on the spliced pRet transcript converting it to a double stranded integrated DNA. Once integrated, the copy in the genome will provide a functional pURA3.

Figure 74 shows the URA3 gene with the mini-intron from the peptide transporter gene inserted in the forward (A) and inverted (B) directions, and the resulting transcripts.

Figure 75 shows the URA3/inverted intron element mounted onto a retrotransposon plasmid in a synthetic Nsi1 site at the 3' end of the coding sequence. An ADE2 element was also inserted between the right LTR and the *Candida* ARS.

Figure 76 shows the retrotransposon plasmid containing a mutated form of URA3 (URΔA3) (A), and a schematic of the URΔA3 and URA3^{+} DNA (B).

Figure 77 schematically represents the results of inverse PCR to analyze and confirm retrotransposition into the genome. --

Please delete the following sections containing drawings in the specification without prejudice, without admission, without surrender of subject matter, and without any intention of creating any estoppel as to equivalents:

- Please delete page 81, line 1 through page 82, line 6.
- Please delete page 82, lines 13-24 and 27-29.
- Please delete page 83, lines 1-5 and 10-20.
- Please delete page 84, lines 13-26.
- Please delete page 86, lines 3-25.